TECHNICAL DATA SHEET

THUNDER™ Total GSPT1 TR-FRET Cell Signaling Assay Kit



CATALOG NUMBERS KIT-GSPT1-100 (100 tests) KIT-GSPT1-500 (500 tests) KIT-GSPT1-2500 (2500 tests) KIT-GSPT1-5000 (5000 tests) KIT-GSPT1-10000 (10000 tests) Store at -80°C For research use only. Not for use in diagnostic procedures.

Visit Product Page

PRODUCT DESCRIPTION

This assay kit measures intracellular levels of total GSPT1 protein in cell lysates using a simple, rapid and sensitive immunoassay based on the homogeneous (no-wash) THUNDER™ TR-FRET technology. The kit is compatible with both adherent and suspension cells.

SPECIFICITY

This assay kit contains two specific and selective antibodies that recognize total GSPT1.

SPECIES REACTIVITY

Human (Swiss-Prot Acc.: P15170-3; Entrez-Gene Id: 2935).

Other species should be tested on a case-by-case basis.

TR-FRET ASSAY PRINCIPLE

The Total GSPT1 assay kit is a homogeneous time-resolved Förster resonance energy transfer (TR-FRET) sandwich immunoassay (Figure 1). The THUNDER™ Cell Signaling assay workflow consists of 3 steps (Figure 2). Following cell treatment, cells are first lysed with the specific Lysis Buffer provided in the kit. Then Total GSPT1 in the cell lysates is detected with a pair of fluorophore-labeled antibodies in a simple "add-incubate-measure" format (single-step reagent addition; no wash steps). One antibody is labeled with a donor fluorophore (Europium chelate: Eu-Abl) and the second with a farred acceptor fluorophore (FR-Ab2). The binding of the two labeled antibodies to distinct epitopes on the target protein takes place in solution and brings the two dyes into close proximity. Excitation of the donor Europium chelate molecules with a flash lamp (320 or 340 nm) or a laser (337 nm) triggers a FRET from the donor to the acceptor molecules, which in turn emit a TR-FRET signal at 665 nm. Residual energy from the Eu chelate generates light at 615 nm. The signal at 665 nm is proportional to the concentration of Total GSPT1 in the cell lysate. Data can be expressed as either the signal at 665 nm or the 665 nm/615 nm ratio.

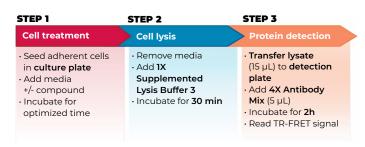


Figure 2 Assay workflow using the 2-plate (transfer) protocol.

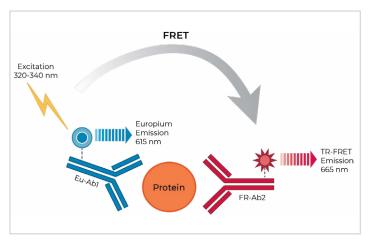


Figure 1 Schematic representation of the TR-FRET cell signaling assay principle.

KIT COMPONENTS	100 points*	500 points*
Eu-labeled total GSPTI antibody (Eu-Abī)	5 μL	25 μL
Acceptor-labeled total GSPTI antibody (FR-Ab2)	20 µL	100 µL
Lysis Buffer 3 (5X)	1 mL	5 mL
Detection Buffer (10X)	50 μL	250 µL
Positive control cell lysate	100 µL	500 μL
Phosphatase Inhibitor Cocktail (100X)	50 μL	250 µL

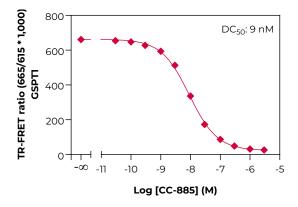
^{*} The number of assay points is based on an assay volume of 20 µL in half-area 96-well or low-volume 384-well assay plates using the kit components at the recommended concentrations (refer to the User Manual).

VALIDATION DATA

This assay kit has been validated for the relative quantification of total GSPT1 in A549 and HeLa cell lysates using the 2-plate assay protocol.

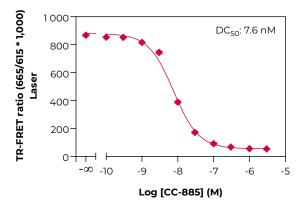
- · Adherent cells were cultured overnight in a 96-well tissue culture plate (RPMI +10% FBS).
- · Following cell treatment, the media was removed and cells were lysed with the 1X Lysis Buffer 3 (50 μ L) supplemented with the 100X Phosphatase Inhibitor Cocktail diluted at 1X.
- · Following a 30-min incubation at room temperature (RT) on
- an orbital shaker (400 rpm), lysates (15 μ L) were transferred to a 384-well assay plate followed by addition of the labeled antibodies Eu-Ab1 and FR-Ab2 (5 μ L) for detection of total GSPT1.
- The plate was incubated at RT for **2 hours** and the TR-FRET signal was recorded at 665 and 620 nm (PHERAstar® *FSX*; laser excitation).

DEGRADATION OF GSPT1 IN A549 CELLS



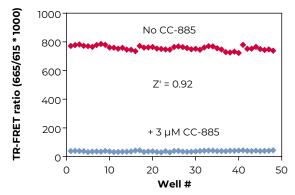
A549 cells (10,000 cells/well; in triplicate) were incubated with serial dilutions of the molecular glue CC-885 for 24 hours at 37°C. Data show that treatment of A549 cells with CC-885 degrades total GSPT1.

DEGRADATION OF GSPT1 IN HELA CELLS



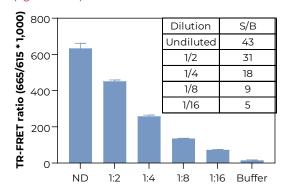
HeLa cells (50,000 cells/well; in triplicate) were incubated with serial dilutions of the molecular glue CC-885 for 24 hours at 37°C. Data show that treatment of HeLa cells with CC-885 degrades total GSPTI.

A549 CONTROL LYSATE TITRATION (QC TEST)



A549 cells (10,000 cells/well) were incubated with or without $3~\mu\text{M}$ of CC-885 for 24 hours at 37°C. The Z' factor value was determined after a 2-hour incubation period, using a total of 48 wells for each treatment group. The Z'-factor value of 0.92 indicates that the assay is robust and suitable for HTS.

A549 CONTROL LYSATE TITRATION (OC TEST)

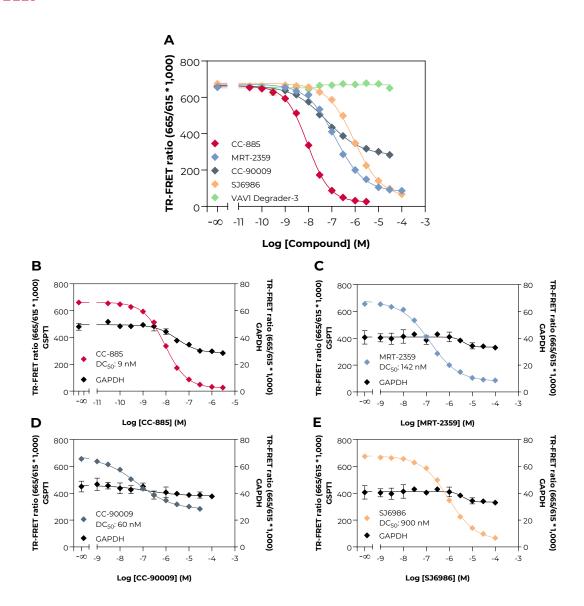


Quality Control: the total GSPT1 assay kit is routinely tested against A549 lysates. A549 cells were cultured in a T175 flask to 85% confluence. Following cell lysis using 4 mL of 1X Lysis Buffer 3, lysates were serially diluted with 1X Lysis Buffer 3 and tested in triplicate. Data show a linear relationship between lysate dilutions and TR-FRET ratio values.



FOR MORE INFORMATION ON DEVELOPING AND OPTIMIZING TR-FRET CELL SIGNALING ASSAYS, CONSULT THE USER MANUAL.

EFFECT OF DIFFERENT MOLECULAR GLUES ON GSPT1 DEGRADATION IN A549 CELLS



A549 cells (10,000 cells/well; in triplicate) were incubated with serial dilutions of molecular glues CC-885, MRT-2359, CC-90009, SJ6986, or VAV1 degrader-3 (as a negative control) for 24 hours at 37°C. Data show that the four molecular glues CC-885, MRT-2359, CC-90009, and SJ6986 triggered a dose-dependent decrease of total GSPT1 in A549 cells, whereas GSPT1 levels were not modulated by VAV1 degrader-3 (A). Treatment of A549 cells with the four molecular glues CC-885, MRT-2359, CC-90009, and SJ6986 degrades total GSPT1 whereas levels of housekeeping total GAPDH (measured as a control) are only slightly affected (B, C, D, E).