

TECHNICAL DATA SHEET



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THUNDER™ Total SLP-76 TR-FRET Cell Signaling Assay Kit

CATALOG NUMBERS KIT-SLP-76T-100 (100 tests)
KIT-SLP-76T-500 (500 tests)

Store at -80°C
For research use only.
Not for use in diagnostic procedures.

PRODUCT DESCRIPTION

This assay kit measures intracellular levels of **total SLP-76** protein in cell lysates using a simple, rapid and sensitive immunoassay based on the homogeneous (no-wash) THUNDER™ TR-FRET technology. The kit is compatible with both adherent and suspension cells.

SPECIFICITY

This assay kit contains two specific and selective antibodies that recognize **total** (both phosphorylated and unphosphorylated) **SLP-76**.

SPECIES REACTIVITY

Human (Swiss-Prot Acc.: Q13094; Entrez-Gene Id: 3937).

Other species should be tested on a case-by-case basis.

TR-FRET ASSAY PRINCIPLE

The **Total SLP-76** assay kit is a homogeneous time-resolved Förster resonance energy transfer (TR-FRET) sandwich immunoassay (Figure 1). The THUNDER™ Cell Signaling assay workflow consists of 3 steps (Figure 2). Following cell treatment, cells are first lysed with the specific Lysis Buffer provided in the kit. Then **Total SLP-76** in the cell lysates is detected with a pair of fluorophore-labeled antibodies in a simple "add-incubate-measure" format (single-step reagent addition; no wash steps). One antibody is labeled with a donor fluorophore (Europium chelate; Eu-Ab1) and the second with a far-red acceptor fluorophore (FR-Ab2). The binding of the two labeled antibodies to distinct epitopes on the target protein takes place in solution and brings the two dyes into close proximity. Excitation of the donor Europium chelate molecules with a flash lamp (320 or 340 nm) or a laser (337 nm) triggers a FRET from the donor to the acceptor molecules, which in turn emit a TR-FRET signal at 665 nm. Residual energy from the Eu chelate generates light at 615 nm. The signal at 665 nm is proportional to the concentration of **Total SLP-76** in the cell lysate. Data can be expressed as either the signal at 665 nm or the 665 nm/615 nm ratio.

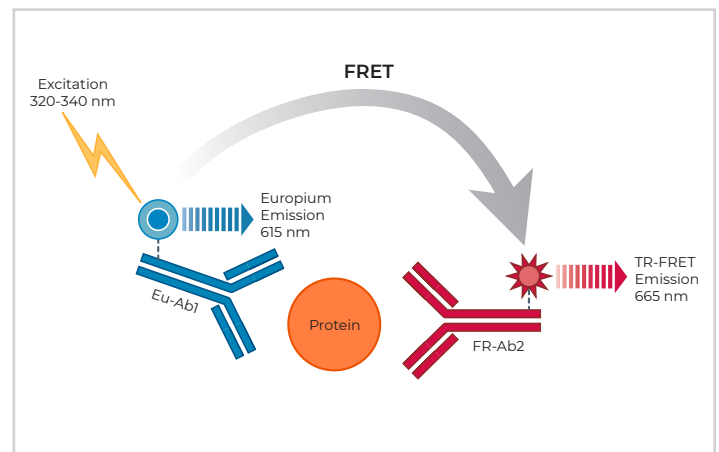


Figure 1 Schematic representation of the TR-FRET cell signaling assay principle.

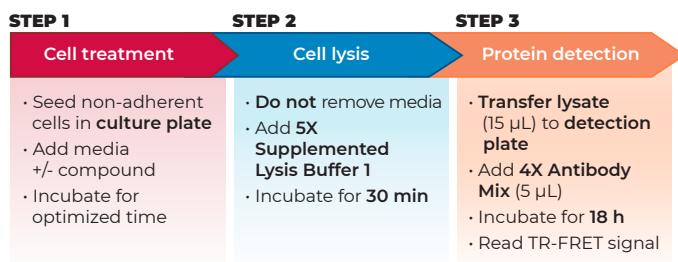


Figure 2 Assay workflow using the 2-plate (transfer) protocol.

KIT COMPONENTS

	100 points*	500 points*
Eu-labeled total-SLP-76 antibody (Eu-Ab1)	5 µL	25 µL
Acceptor-labeled total-SLP-76 antibody (FR-Ab2)	20 µL	100 µL
Lysis Buffer 1 (5X)	1 mL	5 mL
Detection Buffer (10X)	50 µL	250 µL
Positive control cell lysate	100 µL	500 µL

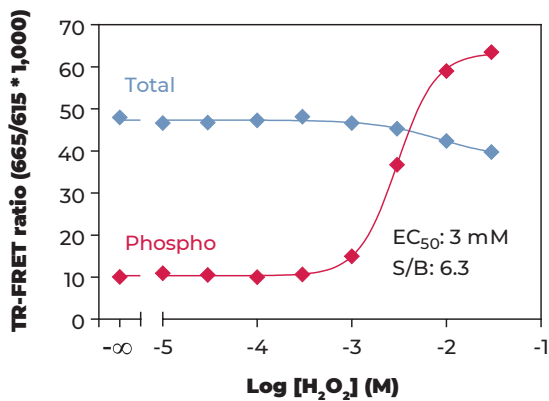
* The number of assay points is based on an assay volume of 20 µL in half-area 96-well or low-volume 384-well assay plates using the kit components at the recommended concentrations (refer to the User Manual).

VALIDATION DATA

This assay kit has been validated for the relative quantification of total SLP-76 in Jurkat cell lysates using the 2 plate assay protocol.

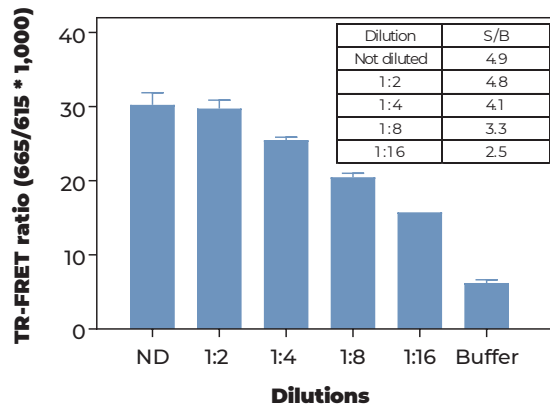
- Non-adherent cells were cultured in RPMI+10% FBS before being centrifuged and resuspended at the desired density in RPMI without serum.
- Following cell treatment, cells were lysed with the 5X **Lysis Buffer 1** (to a final concentration of 1X) supplemented with the phosphatase inhibitors sodium fluoride (5 mM) and sodium orthovanadate (10 mM).
- Following a **30-min** incubation at room temperature (RT) on an orbital shaker (400 rpm), lysates (15 μ L) were then transferred to a 384-well assay plate followed by of the labeled antibodies Eu-Ab1 and FR-Ab2 (5 μ L) for detection of total SLP-76.
- The plate was incubated at RT for **18 hours** and the TR-FRET signal was recorded at 665 and 615 nm (EnVision[®]; lamp excitation).

STIMULATION OF PHOSPHO-SLP-76 (S376) IN JURKAT CELLS



Jurkat cells (400,000 cells/well; in triplicate) were incubated with serial dilutions of H₂O₂ for 15 min at RT. Data show that treatment of Jurkat cells with H₂O₂ stimulates phosphorylation of SLP-76 at S376 but does not affect the levels of total SLP-76.

JURKAT CONTROL LYSATE TITRATION (QC TEST)



Quality Control: the Total SLP-76 assay kit is routinely tested against H₂O₂ treated Jurkat lysates. Jurkat cells were cultured in a T175 flask, centrifuged and resuspended at 20 million cells/mL, and stimulated with 30 mM of H₂O₂ for 15 min at RT. Following cell lysis using 5X Lysis Buffer 1 (final concentration: 1X), lysates were serially diluted with 1X Lysis Buffer 1 and tested in triplicate. Data show a linear relationship between lysate dilutions and TR-FRET ratio values. Note that due to the very high sensitivity of the Total STAT3 kit, lysates from the T175 flask required at least a 1:2 pre-dilution in order to be within the dynamic assay range.

